

Diamond™ Nucleic Acid Dye

INSTRUCTIONS FOR USE OF PRODUCT H1181.

Quick
PROTOCOL

Protocol

Materials to Be Supplied By the User

- plastic staining trays
- dilution buffer (TE, TAE or TBE buffer)

Preparation of 1X Staining Solution

1. Thaw the Diamond™ Nucleic Acid Dye completely at room temperature (22°–25°C) protected from light. Vortex briefly.
2. Prepare a 1:10,000 dilution of the dye in 1X TE, TAE or TBE buffer.

Note: For best results, the dye dilution buffer should be the same as the buffer used to cast the gel. The dilution buffer should have a pH of 7.0–8.5. Do not use water to dilute the dye.

Staining the Gel

1. After electrophoresis, place the gel in a plastic staining tray and completely cover the gel with staining solution.

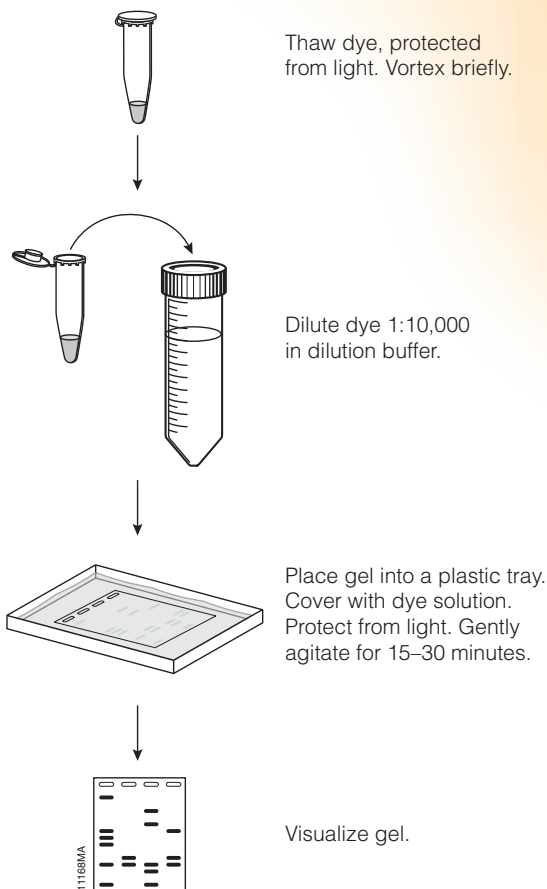
Note: Pipette tip box lids or similarly sized plastic containers make convenient staining trays. We **do not** recommend the use of glass containers for staining.

2. Incubate the gel in staining solution at room temperature (22°–25°C) on a rocker or orbital shaker with gentle agitation, protected from light, for 15–30 minutes. Staining time depends on the size, thickness, and the percentage of agarose or polyacrylamide in the gel.

Visualizing and Documenting the Gel

1. Visualize the gel using a UV epi-illuminator or transilluminator or another gel documentation system, with a maximum excitation of 495nm.
2. Document the gel using Polaroid 667 film or a digital camera with a 500nm cutoff filter, or another gel documentation system that detects emission at 558nm.

Detailed protocols and instructions are available in the *Diamond™ Nucleic Acid Dye Technical Manual #TM388*, available at: www.promega.com/protocols/



ORDERING/TECHNICAL INFORMATION:

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